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Analysis of Haplotype Sequences

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Abstract

In this era of whole-genome, next-generation sequencing, it is important to have a clear understanding of the concept of “haplotype”. We show here that most of the important regions of the genome can be described in terms of polymorphic frozen blocks (PFB). At each PFB, there are numerous, even hundreds, of alternative ancestral haplotypes. Haplotypes, not genes, can be regarded as the principal unit of inheritance. We illustrate how sequence data can be analysed to reveal and define these ancestral haplotypes.

Keywords: Ancestral haplotypes, Polymorphic frozen blocks, Genomic evolution

1. Introduction

Comparative analyses of haplotype sequences allow many efficiencies. It is not surprising that there are many enthusiastic claims. Haplotypes, by any of many definitions, offer opportunities to understand the inheritance of polymorphic traits and their regulation. The most useful are markers of extensive complex polymorphic sequences of evolutionary significance even when the functional components, whether coding or noncoding, are yet to be elaborated.

Substantial advances became possible with the elucidation of genomic structure and function more than 20 years ago and long before recent advances in sequencing technology [1] and bioinformatics [2]. It became clear that haplotypes, *not genes*, can be regarded as the principal unit of inheritance.

This chapter evaluates some competing strategies and illustrates the power now available through NGS.

2. Haplotype terminology

A review of current literature reveals a staggering collection of terms synonymous with haplotypes, as listed in Table 1.

Ancestral haplotypes
Conserved extended haplotypes
Linkage groups
Linkage disequilibrium haplotypes
Hapmaps
Haplogroup
Haplobanks
Haploblocks
Haplotype block

Table 1. Terminology

Even if it were possible to define the various neologisms, it seems certain that confusion will remain until there is recognition of the conceptual background.

We introduced the term *ancestral haplotypes* to emphasise the persistence of the founding pool [3, 4]. Such haplotypes are conserved over thousands of generations; they allow identification of remote ancestors and their contributions to the creation of individual members of the species with their diseases. Unfortunately, others use the same term in different ways and even in the opposite sense, that is, to refer to *the single* original haplotype which is presumed to have mutated to give rise to all the so-called variants now present. Indeed, as just one example of the problem, the reader has to be able to interpret the following: "we identified all nonredundant haplotypes with a frequency of $\geq 10\%$ and consisting of at least 10 SNPs, which are likely to represent the nonrecombinant descendants from a single ancestor" [5].

To yet further confound matters, increasingly, the term *haplotype* is being used to describe any combination of alleles or markers, such as SNPs, without regard to their reproducibility, inheritance, polymorphism or biological significance. Currently, there are conflicting methods of detection. The problems appear to be increasing as ephemeral concepts diverge and as claims for better approaches focus on just one or another competing technology or bioinformatic package.

Several other aspects are clear.

- Linkage groups relate to closely linked loci but do not define haplotypes.
- Linkage disequilibrium is affected by relative frequencies and therefore fails to detect rare haplotypes.
- Trios can be misleading since the coverage of the family is limited.
- Haplobanks. The Tokunaga group has established some important principles with the intention of establishing haplotype-matched pluripotential stem cell banks [6]. Unfortu-

nately, and amazingly, there is now uncertainty as to how to define the haplotypes. For example, a recent paper urges international collaboration to avoid fragmentation [7]. It would be wise to avoid neologisms and such redefinitions without clarity of meaning.

3. Definitions and concepts

In the presequencing era, there was a clear understanding of what was meant by the term *haplotype*: Combinations of alleles at different loci segregating together in multigenerational family studies [8]. Some seem unaware of this long history and have had to rediscover the concept [2].

The implications were apparent at least 50 years ago: a specific allele A1 at locus A is inherited together with a specific allele B1 at an adjacent, “closely linked” locus B [9]. The fact that these two alleles segregated together through multiple generations was unexpected and led to controversy but, in retrospect, clearly implied that

1. The two alleles were encoded on the same chromosome, whether paternal or maternal.
2. The two loci were closely linked.
3. Recombination was rare.
4. The two loci arose by duplication.
5. Duplication is associated with polymorphism.

The repeated cosegregation of alleles came to be known as a haplotype: from ἀπλφούς = single [9].

It is worth emphasizing that it was the cosegregation as haplotypes through “phased” multigenerational families (rather than “unphased” populations) which foretold the later demonstration that there was a continuous haplospecific sequence. It is also pertinent, with the benefit of hindsight and in view of recent confusion, that the haplotypes, defined in one family, occurred in other families of similar remote ancestry raising the radical possibility of conservation beyond that expected from close linkage alone. In other words, recombination is patchy and does not necessarily disperse the components of duplications, even after thousands of meioses. The issue of linkage disequilibrium and the limits of LD mapping are considered below.

The implications of haplotypes, as listed above, became even clearer as the HLA A and HLA B locus alleles and then HLA DR alleles were defined during the 1970s. However, in this case, the loci were widely separated. Over time, it became clear that each of the A-B and B-DR haplotypes were some 800 kb in length. Patently, close linkage could not explain these haplotypes; either there was selection for *cis* interaction or there was suppression of recombination [3, 4].

Through their studies of diseases, the Alper–Yunis group discovered that the B-DR haplotypes contained specific alleles at duplicated loci which had no structural or functional relevance to HLA (i.e. complement and 21 hydroxylase loci) but which happen to be located within the

major histocompatibility complex [10–16]. Thus, *cis* interaction alone could be rejected as the sole explanation.

The importance of discovery through disease was illustrated at a meeting held in 1982 [3, 4]. As shown in Table 2, it was disease associations which allowed the initial discovery of ancestral haplotypes; note, these three disease-associated haplotypes could have only been discovered through their associations. Two share DR3 and two share B18 but the frequencies differ. Thus, the three haplotypes cannot be detected by linkage disequilibrium.

Designation	A	Cw	B	Bf	C2	C4A	C4B	DR	Disease
8.1	1	7	8	S	C	Q0	1	3	MG, SLE, IDDM
18.2	–	–	18	F1	C	3	Q0	3	IDDM
18.1	25	–	18	S	Q0	4	2	2	C2 deficiency

MG = myasthenia gravis, SLE = systemic lupus erythematosus, IDDM = insulin-dependent (type 1) diabetes mellitus.

Adapted from ref. [4]

Table 2. MHC haplotypes and disease associations

Once the numerous other ancestral haplotypes were defined, multigenerational family studies identified cosegregating combinations of multiple alleles at separated loci, i.e. haplotypes stretching over nearly 2 Mb from HLA A to DR. A haplotype was defined by the alleles “inherited *en bloc* from one parent and implies the transmission of all of the chromosomal segment” from one generation to the next [4].

When haplotypes defined in one family were compared with those identified in apparently unrelated families, sharing was immediately apparent. There were specific combinations of alleles at all the numerous unrelated loci as these were defined and typed. However, and increasingly relevant today, as summarized in refs. [3, 4, 17, 18]:

1. The combinations observed are *not* a simple function of allele frequencies; only some of the components inherited *en bloc* are in linkage disequilibrium.
2. Many haplotypes are rare combinations of frequent alleles at some loci but rare alleles at other loci.
3. Very few alleles are entirely haplospecific.
4. Haplotype frequencies are often less than 1%.
5. The same haplotypes are found in multiple, apparently unrelated, families.
6. Many of these nonrandom combinations are associated with a disease (such as systemic lupus erythematosus) or function (such as TNF production).
7. With a few dramatic exceptions (such as 21 hydroxylase and C2 deficiency carried by what we now call the 47.1 and 18.1 ancestral haplotypes), the individual alleles do not explain the haplospecific effects on disease and function.

8. Penetrance is low. That is to say, the haplotypes are *sine qua non* in that they permit particular diseases and functions but only in the presence of other genetic, infectious, environmental, hormonal and age-related factors.
9. Recombination is rare and difficult to demonstrate even within multigenerational families with the potential to confirm a meiotic recombinant. Nevertheless, over the life of an ancestral haplotype—say 10, 000 meioses—there have been recombinations which have resulted in shuffling between ancestral haplotypes [18, 19].

[illegible]

Adapted from ref. [18].

Figure 1. Historic recombinations of AH 8.1. The HLA-B8 allele is carried by one ancestral haplotype marked by A1, Cw7, B8, BfS, C4AQ0, C4B1, DR3. All the haplotypes in data set 1 carrying HLA-B8 are represented. These haplotypes have been sorted so that haplotypes that carry all alleles of 8.1 from HLA-A to DR are shown at the top of the figure, followed by haplotypes that extend from HLA-B to DR. Telomeric recombinants are shown at the bottom. The boxed areas represent those portions of the 8.1 ancestral haplotype that are carried by unrelated B8-containing haplotypes. Vertical lines approximately indicate the region where historical recombination has occurred.

Some of these points are illustrated in Figure 1. It can be seen that subjects with B8 can be listed to show conservation but also historic recombinations between HLA A and B, between C4B and DR, and between HLA B and Bf.

By the mid-1990s, and long before the rediscoveries of the 2000s [2], such analyses led to the conclusion that there are polymorphic frozen blocks (PFB), as illustrated in Figure 2.

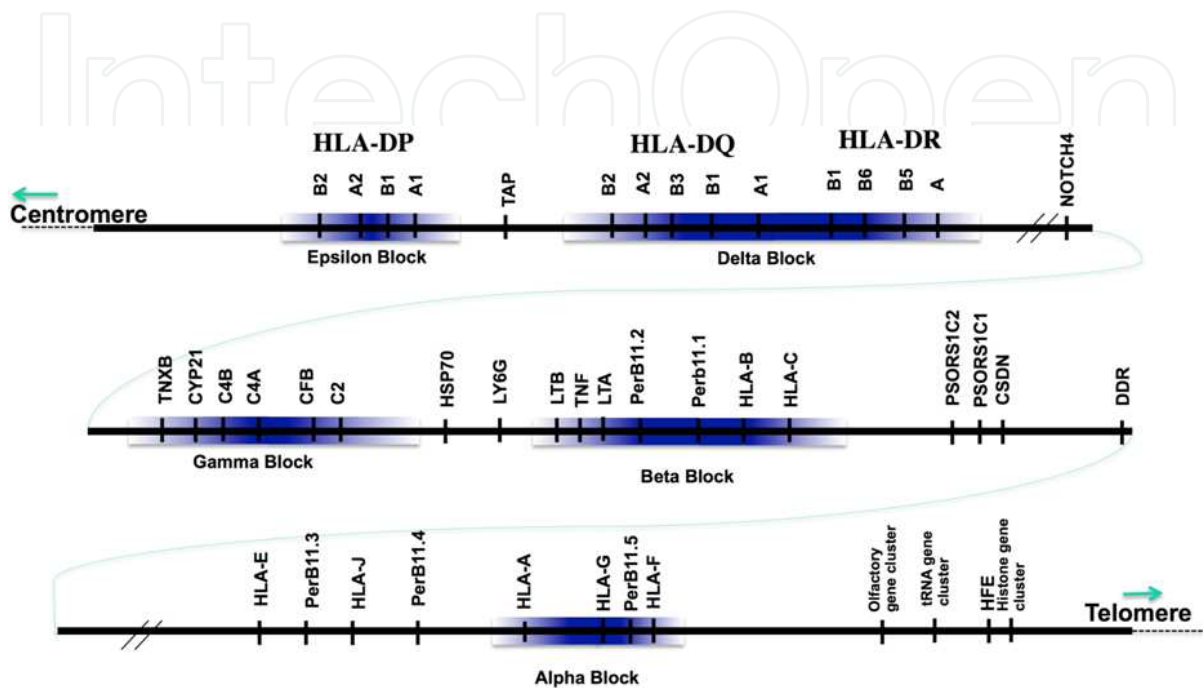
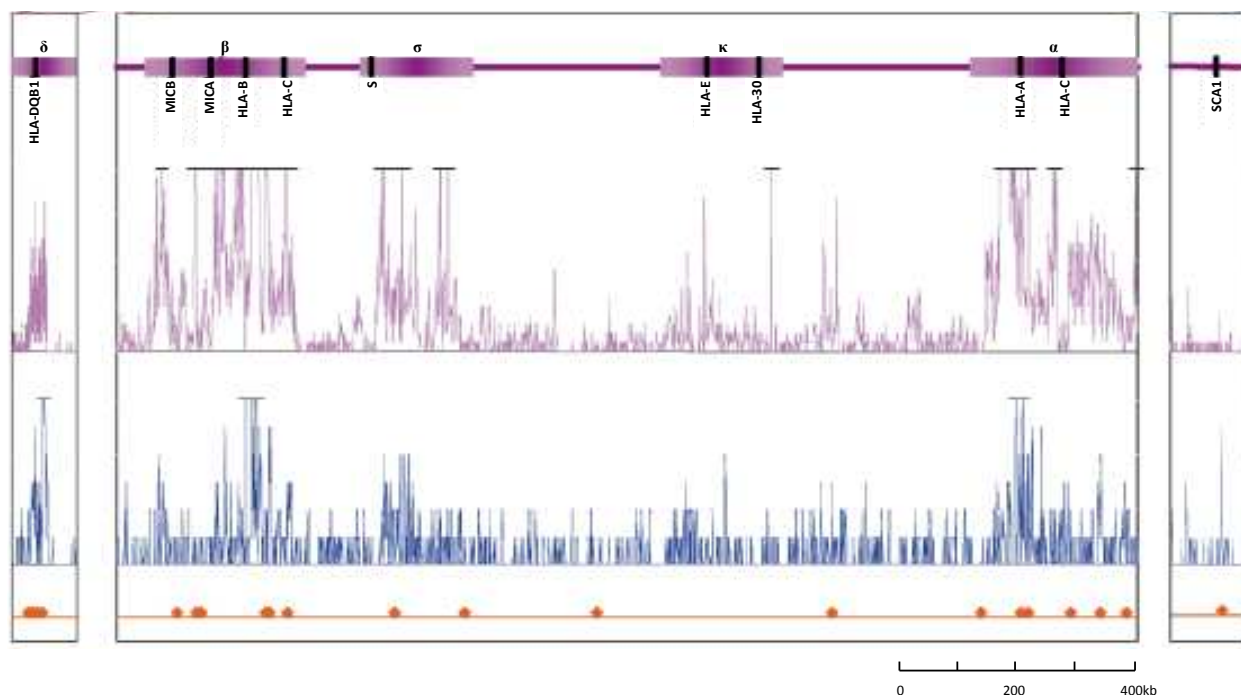


Figure 2. Ancestral haplotypes and polymorphic frozen blocks within the human major histocompatibility complex. Each ancestral haplotype has its own unique DNA sequence which includes single nucleotide polymorphisms (SNPs), copy number variations, segmental duplications, insertion and deletion events (indels) including retroviral and retroviral-like elements (RLEs). The full length is approximately 4 Mb. Higher degrees of diversity indicated by shading define polymorphic frozen blocks (PFB). Recombination occurs far more frequently between, rather than within, these blocks. Mutations within blocks are effectively suppressed. Adapted from refs. [17, 20] and [21]. Reproduced with permission from ref. [22].

PFB throughout the genome are the latter-day equivalents of loci. Sequences which define ancestral haplotypes are the equivalent of alleles. The diversity is multifactorial with contributions from reiterative speciation as follows [17]:

- Retroviral integration
- Duplication
- Indels
- Polymorphism

These elements all contribute to the haplospecificity of the sequence of ancestral haplotypes as shown in Figure 3. Similar distribution of diversity has been found by many others [5, 17, 19, 20, 23, 24]. The same patterns are also found in primates [25].



Adapted from ref. [26].

Figure 3. Sequence diversity is packaged as polymorphic frozen blocks (PFB). SNPs and indel occur in similar locations within PFB. (a) The SNP profile after removing indels. Peaks higher than 20 SNPs per 1000 nucleotides are truncated. (b) The location of indels. Peaks higher than six indels per 1000 nucleotides are truncated. (c) The position of indels greater than 100 nucleotides.

4. Use of ancestral haplotypes

Here, we illustrate the potential of sequence analysis, if designed to identify conserved, extended, ancestral haplotypes. The utility depends very largely on the concept behind the analysis. However, it also depends upon the genomic region actually sequenced and whether it is possible to interpret the patterns in the context of the heterogeneous architecture of the genome. Within PFB, there will be a multitude of alternative sequences to compare. In the genome between these blocks, there is much less diversity with long stretches of monomorphic sequence. Thus, the recent fashion for identifying homozygosity [27, 28], without regard to diversity, shifts the focus to less informative regions of the genome. Of course, by way of explanation for the fashion, homozygosity within PFB is much more difficult to find; the most common ancestral haplotypes with frequencies of 0.1 will be homozygous in only 1% of the general population. Until high-throughput NGS became available, it was necessary to examine disease panels or consanguineous families.

The conceptual background is summarised in the following figures which contrast two approaches. *Population genetics* teaches that free recombination effectively prevents the packaging of polymorphism. The reality, designated here as *quantal genomics*, emphasises clustering and conservation of polymorphism. Each haplotype is a specific sequence which

regulates expressed genes by *cis*, *trans* or *epistatic* interaction. The whole sequence is conserved. Linkage disequilibrium, when it occurs, is simply a reflection of this conservation which includes haplotypes with alleles which are relatively common in one haplotype when compared with others. Each is ancestral, in the sense that they are shared by apparently unrelated families separated by hundreds or even thousands of generations. It follows that the polymorphisms are actively conserved and could not be a consequence of recent mutation.

Some of the implications are illustrated in Figures 4 and 5.

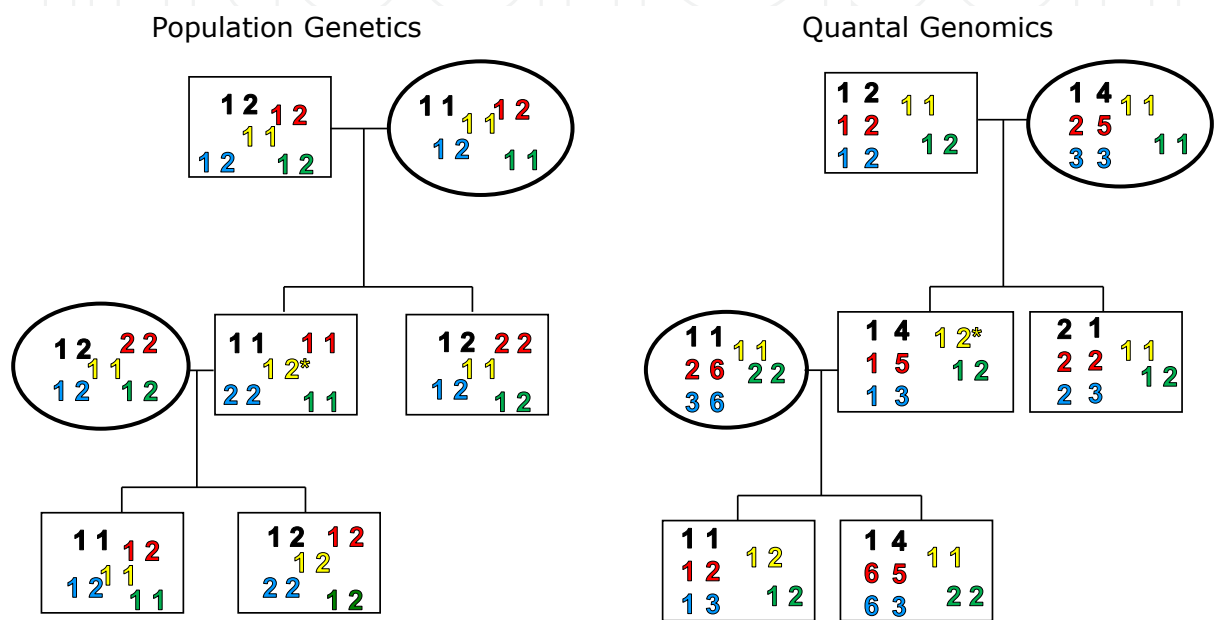


Figure 4. Importance of clustering functional genes. Colours represent loci and numbers represent alleles at those loci. On the left is the basis of the infinitesimal model used in population genetics. Loci are biallelic and can be homozygous or heterozygous. Free recombination occurs between loci and alleles segregate independently. On the right, loci are within polymorphic frozen blocks (PFB), shown by alignment of loci. Alleles within PFB segregate *en bloc*, forming haplotypes, which are inherited intact through many generations. Important genes are carried within PFB, conserving their *cis* interactions. Loci within PFB have multiple alleles, allowing for a greater degree of polymorphism clustered within the block. There can be hundreds of ancestral haplotypes for each PFB. *Trans* interactions between haplotypes increase the diversity expressed in the population. The loci shown in green and yellow are outside the PFB and follow a pattern of inheritance similar to population genetics. *De novo* mutations are indicated by asterisk—on the right the mutations occur at loci outside of conserved PFB and will have little if any consequence because truly important differences are encoded within PFB. Monogenic diseases or traits are the partial exceptions. On the left, mutations can occur at any loci but are generally assumed to occur at loci that were monoallelic. They may or may not be important, depending upon frequency, context, repair and heritability. Adapted with permission from ref. [22].

By 1987, it was clearly established that each ancestral haplotype has a specific content of genomic features such as duplications and indels. These too are actively conserved and can themselves be used as signatures for haplotypes of hundreds of kilobases and even megabases. These observations were very difficult to explain in terms of any form of neo-Darwinism, natural selection, random errors or population genetics as taught then and today. Rather, we realised, the genome is not actually homogeneous but partitioned into protected quanta or PFB [17, 22, 26, 29].

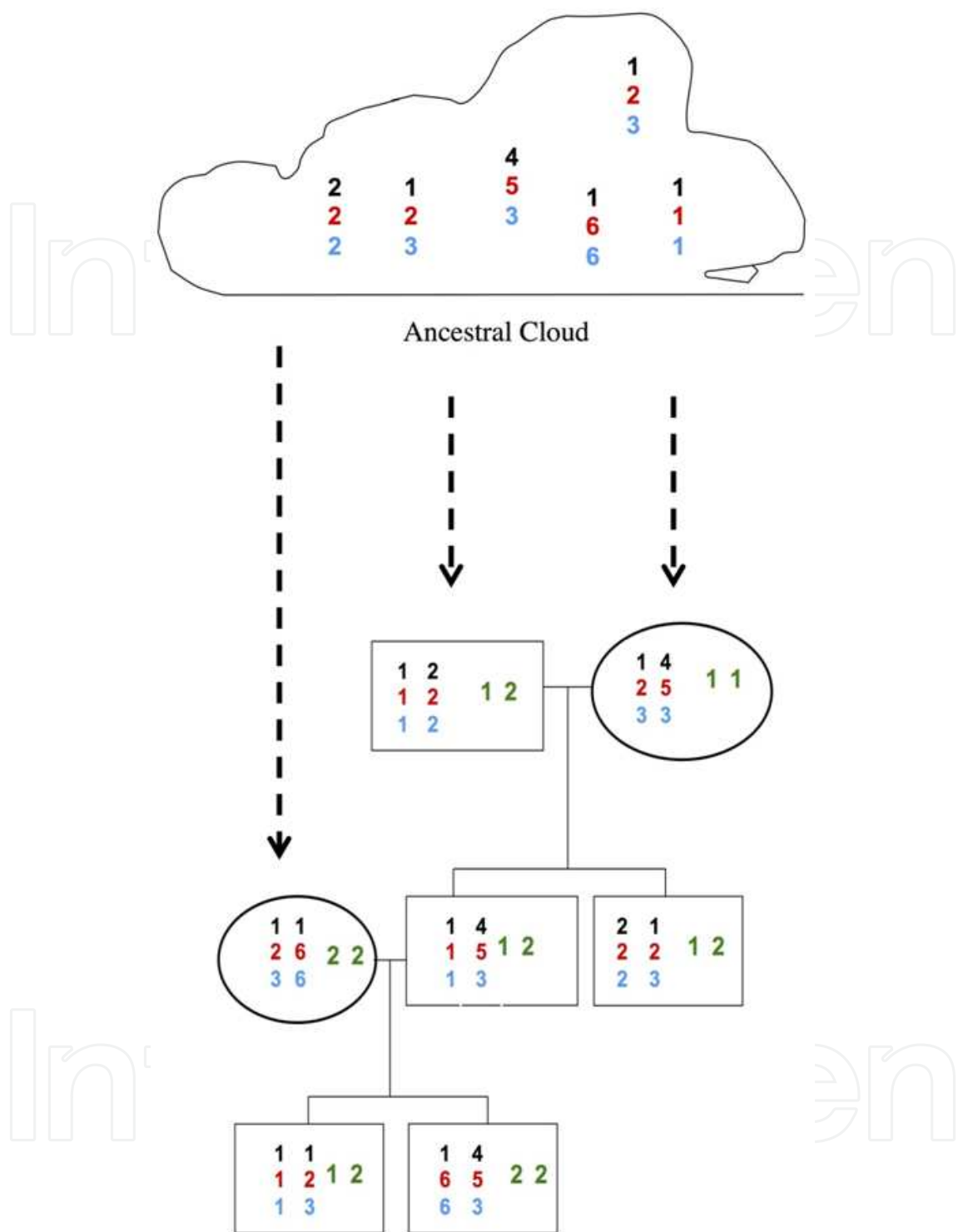


Figure 5. Modern haplotypes are derived from the deep past—they are ancestral haplotypes.

5. Sequencing of critical genomic regions

By 1992, there was sufficient sequencing to confirm the earlier prediction that each ancestral haplotype is actually a frozen sequence.

Haplotype	Geometric element at CL1	Length	Geometric element at CL2	Length
57.1	(TC) ¹² (TG) ⁶ (TC) ¹⁴ (TG) ³ (TC) ¹²	94	TA (TC) ¹⁸ TT (TC) ⁹	58
18.2	(TC) ¹⁴	28	Deleted	
8.1	(TC) ²⁸	56	(TC) ¹⁵ TG (TC) ⁶ TG (TC) ⁸ TG (TC) ⁵	96
7.1	(TC) ¹² (TG) ⁶ (TC) ¹⁴ (TG) ³ (TC) ¹²	94	(TC) ¹⁴ TG (TC) ⁶ TG (TC) ⁸ TG (TC) ⁵	94

Adapted from ref. [30].

Table 3. Haplospecific geometric elements. Ancestral haplotypes have specific sequence signatures at each of the duplicons. Note in 18.2, the duplication did not occur or has been deleted.

We now know that examples of the 8.1 ancestral haplotype are almost identical over megabases [31, 32].

We illustrate the differences between different haplotype sequences in Figure 6. It can be seen that there are certain sites where haplotypes differ. Importantly, haplospecificity is conferred by the whole sequence rather than single nucleotide polymorphisms. For example, reading from left to right, 8.1 and 18.2 differ in T/G but not A/G, etc. Note also that some of the differences are due to indels. Of critical importance is accurate, unmolested sequencing over kilobases, as is now possible through NGS. It is clear, however, that assembly is hazardous especially in areas of duplication and polymorphism. Note also, that there is no justification for regarding *one* particular sequence as the reference. Rather, it is necessary to compare each output with a library of known sequences within each PFB.

The number of differences depends on which haplotypes are compared (see Table 4). Two of the most common Caucasian haplotypes, 8.1 and 7.1, differ by a hundred positions, representing approximately 1% nucleotide diversity. The most different haplotypes are 18.2 and 7.1, having 2.5% nucleotide diversity. Interestingly, these haplotypes are different functionally; 18.2 permits insulin-dependent diabetes mellitus whereas 7.1 is protective.

AH Haplotype	44.2	62.1	7.1	44.1*	8.1
44.2	0				
62.1	187	0			
7.1	249	221	0		
44.1*	73	154	227	0	
8.1	224	219	101	204	0
18.2*	184	130	250	137	245

Table 4. Pairwise differences between haplotypes. Total differences between each pair of haplotypes in the 9277 bp region at HLA-B.

44.2 60 CAGATG 39 AGGACCAGGGT G 16 TGGTGT 14 AGAGGCAG 11 AGGGA 29 TCTTGG 17 TGGTTCTGTGGCC 29 CA- ATATACAACCTTTATG 13 AACTTG 41
62.1 // . . . G . . // . . . C A . . // . . . C . . // . . . G // // // //
7.1 // . . . T G . . // . . . C A . . // . . . C . . // . . . G // // // //
44.1* // . . . T G . . // . . . C A . . // . . . C . . // . . . G // // // //
8.1 // . . . T G . . // // . . . C . . // . . . G . . T . . // // . . . C C . . // . . . C A . . // . . . G G T G . . //
18.2* // . . . G . . // . . . C A . . // . . . C . . // . . . G // . . . A . . // // // //

44.2 TGTGGGC - - GT 55 CTGCC 62 CATCC 31 TTAAACAGAGT 55 GGTTCCCAATCT 9 ATAGG 6 GCGGG 31 AACCC 9 AGTATTATGC 37 GGTGA 12 GATGTG
62.1 . . C GA . . // . . . A . . // . . . G . . // . . . C G . . // . . . C // // //
7.1 . . C GA . . // . . . A . . // . . . G . . // . . . C G . . // . . . C // // //
44.1* . . C GA . . // . . . A . . // . . . G . . // . . . C G . . // . . . C T . . // // //
8.1 . . C TGA . . // . . . A . . // . . . G . . // . . . C G . . // . . . C // . . . G . . // // . . . T . . //
18.2* GA . . // // . . . G . . // . . . C G . . // . . . C T . . // // //

44.2 9 GGTGCTC 42 CAGCC 25 TCGAC 9 CCAGG 17 AGATG 16 GGGTT 21 ACGTGGGGCC 25 TGGTC 19 TGGAGGT 36 TCAGT 7 TTTGTGCCTCATCCGTGCTTG
62.1 // T . . // // // . . . A . . // // . . . T . . // . . . A . . // //
7.1 // T . . // // . . . A . . // . . . T . . // . . . G . . // // . . . A . . // //
44.1* // T . . // // . . . A . . // . . . T . . // . . . G . . // // . . . T . . // //
8.1 // . . . A // . . . A . . // . . . A . . // . . . T . . // . . . G . . // // . . . A . . // //
18.2* // T . . // // . . . A . . // // // . . . T . . // // // . . . G . . //

44.2 11 CTCAT 12 ACTTC 32 CTGAA 27 CCCAGCT 5 ACTTGCCCTTCTCTGGT 16 GAATG 7 TGCCCCCTCCTC 45 CACCC 12 ATCTT 21 TCGTC 7 CACCA 60 ACGTCC
62.1 // // . . . C . . // . . . A . . // . . . AG - . . // . . . CC . . . C . . . A . . // . . . G . . // . . . T T . . // . . . T . . //
7.1 // // . . . C . . // . . . A . . // . . . AG - . . // . . . C A . . // . . . G . . // // . . . T . . // . . . T . . //
44.1* // // . . . C . . // . . . A . . // . . . AG - . . // . . . CC A . . // . . . G . . // // . . . T . . // . . . T . . //
8.1 // . . . G . . // . . . C . . // . . . A . . // . . . AG - . . // . . . CC A . . // . . . G . . // // . . . T . . // . . . T . . // . . . A . .
18.2* // // . . . C . . // . . . A . . // . . . AG - . . // . . . CC A . . // . . . G . . // // . . . T . . // . . . T . . //

44.2 11 CACGC 9 GGTAA 27 CTTGA 20 GCTTCCCTCATCCCTCACC 22 GCAGT 24 GTTAT 6 GCCCT 47 GGTGAGATGCA 31 CCGCA 24 TCTGT 19 TGCTC 11 CAG - - A C
62.1 // // // // . . . C G T G . . // . . . C . . // . . . C . . // . . . T . . // . . . C
7.1 // // . . . A . . // . . . C . . // . . . C G G . . // . . . C . . // . . . C . . // . . . T . . // . . . C
44.1* // . . . T . . // . . . A . . // . . . C . . // . . . C G G . . // . . . C . . // . . . C . . // . . . T . . // . . . C
8.1 // // // // . . . C G G . . // . . . C . . // . . . C . . // . . . T . . // . . . C
18.2* // . . . T . . // . . . A . . // . . . C . . // . . . C G G . . // . . . C . . // . . . C . . // . . . T . . // . . . C

44.2 16 GATAT 7 TCCCT 28 AAAAA 118TAGTG 34 GCTAC 18 CTGATTT 156CTGCA 50 AGTGC 31 TCTAC 83 GATGG 51 TCATGA 54 AGAAG 28 AGGGC 48 CGTGT
62.1 // . . . C . . // . . . T . . // . . . C . . // // . . . C . . // . . . - - - . . // // // . . . C . . // . . . G . . // . . . G . .
7.1 // . . . C . . // . . . T . . // . . . C . . // . . . A . . // // . . . - - - . . // . . . A . . // // . . . G . . // . . . C . . // . . . A . .
44.1* // . . . C . . // . . . T . . // . . . C . . // // // . . . - - - . . // . . . A . . // // // // . . . C . .
8.1 // . . . C . . // . . . T . . // . . . C . . // // // . . . - - - . . // . . . A . . // // // . . . C . . // . . . C . .
18.2* // . . . C . . // . . . T . . // . . . C . . // // // . . . - - - . . // . . . A . . // // // . . . G . . // . . . G . .

44.2 7 CCGCCCTGG 30 CCGCT 55 GACG 50 CCGTC 98 CTCTC 121ACATA 44 CAGGA 21 CAAGT 19 ACGGTGGACACGGGGGTGGGC 120TCAGT 69 AGTGA 7 CTAAA
62.1 // . . . A // . . . A . . // // . . . A . . // . . . T . . // // . . . A . . // . . . GA . . // . . . A AA
7.1 // // . . . A . . // // . . . G . . // . . . A . . // // . . . G . . // . . . A . . // // . . . AA
44.1* // // . . . A . . // . . . A . . // . . . A . . // // // // // //
8.1 // // . . . A . . // . . . A . . // . . . G . . // // // // . . . A . . // . . . G . . // . . . A . .
18.2* // // // // . . . G . . // // // // . . . A . . // . . . G . . //

44.2 6 ACCAC 26 TCTCC 17 CCACTGCCCCACCCACCCCCAGACCTGCCACCCACC 69 TGCCT 87 CCGCCCCCATCA 52 TGCC 6 GGGAC 12 GGGCC 21 CTCGT
62.1 // . G. // . C. // . T. // . C. // . G. // . A. // . C. // . T. // . C. // . A. // .
7.1 // . . // . C. // . T. // . C. // . A. // . C. // . T. // . C. // . A. // .
44.1* // . . // . . // . . // . . // . . // . . // . . // . . // . . // . .
8.1 // . . // . . // . . // . . // . . // . . // . . // . . // . . // . .
18.2* // . . // . . // . T. // . CT. // . C. // . C. // . G. // . G. // . A. // . C. // . T. // . C. // . A. // .

44.2 4 GACCC 32 CAAGG 124 CACCAAGGTGA 4 TCCGTC 57 TGCCT 11 TGGATGCTGCTTC 93 GAGCT 18 AGCAG 64 GACGG 11 GACGG 11 GAAGG 28 GG - - - - -
62.1 // . A. // . G. // . T. // . C. // . T. // . C. // . T. // . T. // . T. // . T. // . T. // . G. // . GCTCC A
7.1 // . . // . G. // . T. // . A. // . C. // . A. // . T. // . T. // . T. // . T. // . G. // . - - - - -
44.1* // . . // . . // . . // . . // . . // . . // . . // . . // . . // . . // . . // . .
8.1 // . . // . . // . . // . . // . . // . . // . . // . . // . . // . . // . . // . .
18.2* // . A. // . . // . T. // . . // . T. // . . // . T. // . . // . T. // . T. // . T. // . G. // . GCTCC A

44.2 - - - - - GC 16 AAGAG 163 TCGGT 58 AAGCC 20 TGACC 27 ACATG 51 CTCTCATGGGAC 80 GCCTGGACGC 15 TCTCCTA 34 AGGGA 105 CACGG 39 CAGTATTC T
62.1 GAAGG. // . C. // . . // . . // . . // . G. // . . // . . // . . // . . // . . // . . // . C. // . C.
7.1 - - - - - // . . // . A. // . A. // . G. // . G. // . T. // . . // . . // . . // . . // . . // . .
44.1* - - - - - // . . // . . // . . // . . // . . // . . // . . // . . // . . // . . // . . // . .
8.1 - - - - - // . . // . A. // . . // . G. // . G. // . C. // . G. // . T. // . . // . . // . . // . . // . .
18.2* GAAGG. // . . // . . // . . // . G. // . . // . . // . . // . . // . - - - - - // . A. // . T. // . .

44.2 36 ATCCC 6 GTCCT 57 AAGGG 102 CCCGCGCGCTGCAGCGTCTC 15 GTATC 6 GCGAC 7 ACAGGC 14 CTCAGCT 6 CCACA 36 GCGGT 4 GCCGC 9 GCTCA
62.1 // . . // . . // . . // . . // . . // . . // . . // . . // . . // . . // . . // . . // . . // . .
7.1 // . T. // . - - - - - // . C. // . A. // . C. // . T. G. // . . // . . // . C. // . T. C. // . C. // . T. // . C. // . G. // . G.
44.1* // . . // . . // . . // . . // . . // . . // . . // . . // . . // . . // . . // . . // . .
8.1 // . . // . - - - - - // . . // . C. // . G. // . C. // . G. // . GT. // . GTC. // . . // . . // . . // . G.
18.2* // . . // . . // . . // . . // . . // . . // . . // . . // . G. // . C. // . GT. // . . // . . // . . // .

44.2 36 CGTCCTGGTCATACC 41 ATCCTCTGGATGATG 12 CCCGG 13 CCCGTCCCC - GA 91 GTCTG 16 TCGGGG - GC 36 GTACG 28 CCGGG 36 GCGGAGCGCG G
62.1 // . GA. // . G. // . . // . G. G. // . A. // . . // . T. // . A. // . GG. // . . // . A. // . C. C. AG. T
7.1 // . A. // . G. // . G. // . G. // . A. // . - - - - - // . A. // . AG. // . . // . . // . C. C. AG. T
44.1* // . . // . . // . . // . . // . . // . . // . . // . . // . . // . . // . . // . . // . .
8.1 // . A. // . T. G. // . G. // . G. G. // . . // . . // . C. // . . // . A. // . AG. // . C. // . . // . C. C. AG. T
18.2* // . GA. // . G. // . G. G. // . . // . . // . C. // . . // . A. // . GG. // . . // . . // . C. C. AG. T

44.2 T GCGCAGGTTCTCTCGGTAAGTCTGTGTGTTGGTCTTGGAG 6 GTCTCCC 20 TCCTG 7 CATGG 4 CGCGGCTCCTTCCT 6 CGTGG 21 ACAGCGTGTCTG 12
62.1 . C. // . C. // . . // . . // . . // . . // . . // . . // . G. // . G. // . A. // . . // . C. // . T. G. // .
7.1 . C. // . C. // . C. // . CC. G. C. // . T. // . G. T. // . . // . C. // . . // . CT. // . C. // . T. G. // .
44.1* . . // . . // . . // . . // . . // . . // . . // . . // . . // . . // . . // . . // . .
8.1 . C. // . C. // . C. // . A. // . G. T. // . . // . C. // . . // . CT. // . C. // . T. G. // .
18.2* . C. // . C. // . . // . . // . . // . . // . G. T. // . T. // . C. // . G. // . . // . C. // . T. G. // . C. // .

44.2 ACGGTGA 31 CATGGCGGTGTAGA 24 GGGGCGAG 40 CCCC GG 25 TCTCTCCTCCCCACA 12 CTCCCGACCCCGCACTCACCGGC 18 CACTGCCCCCCAG
62.1 . T. C. // . . // . . // . . // . . // . . // . . // . G. // . G. // . . // . GG. T. // . G.
7.1 . T. A. // . C. A. // . . // . T. // . T. A. // . G. // . GG. // . . // . GG. // . G. G.
44.1* . . // . . // . . // . . // . . // . . // . . // . G. // . GG. // . . // . GG. // . G. G.
8.1 . T. A. // . . // . C. // . . // . T. A. // . G. // . GG. // . - - - - - // . GG. // . G. G.
18.2* . T. A. // . C. A. // . G. // . A. // . . // . G. // . . // . G. // . . // . A. // . .

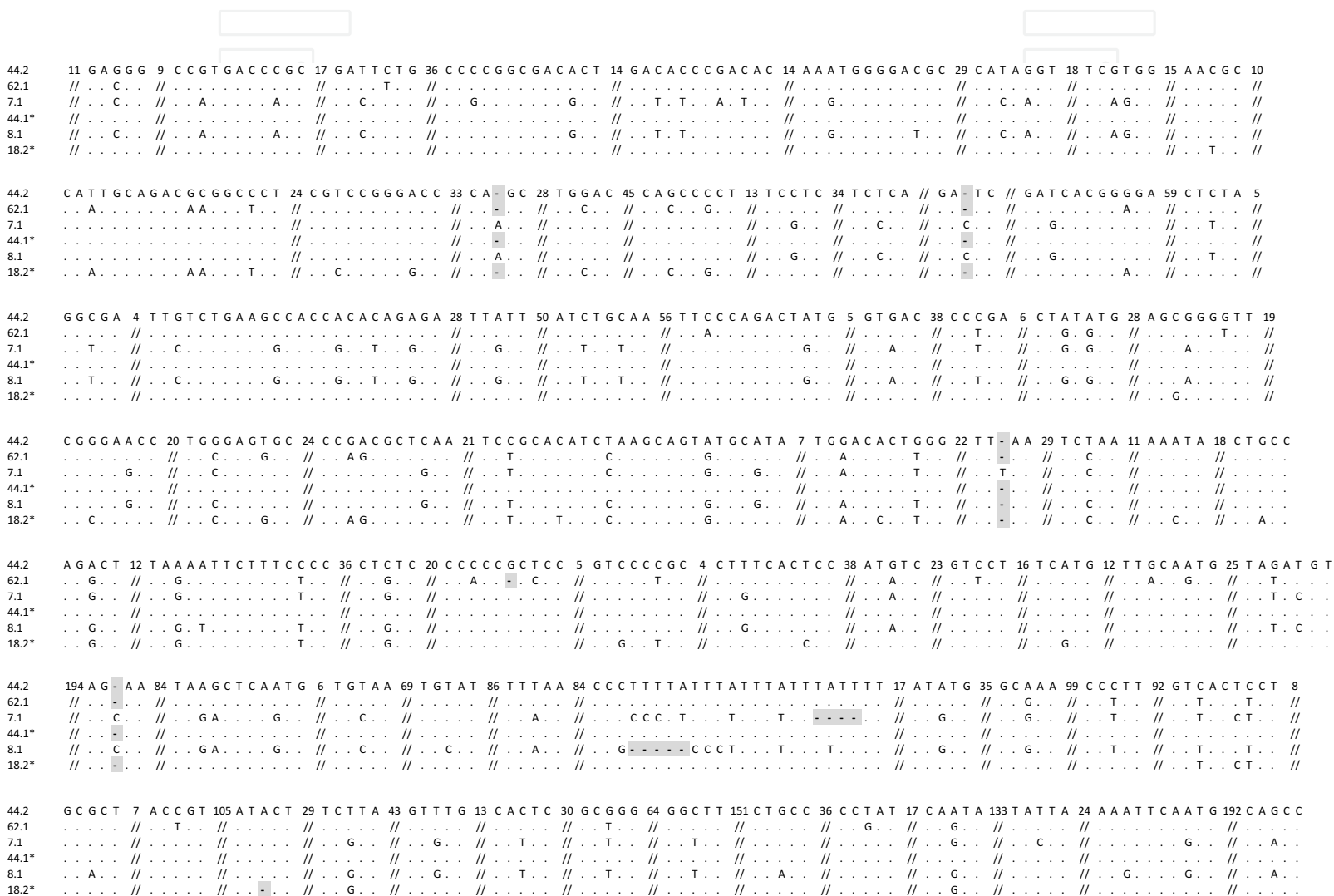
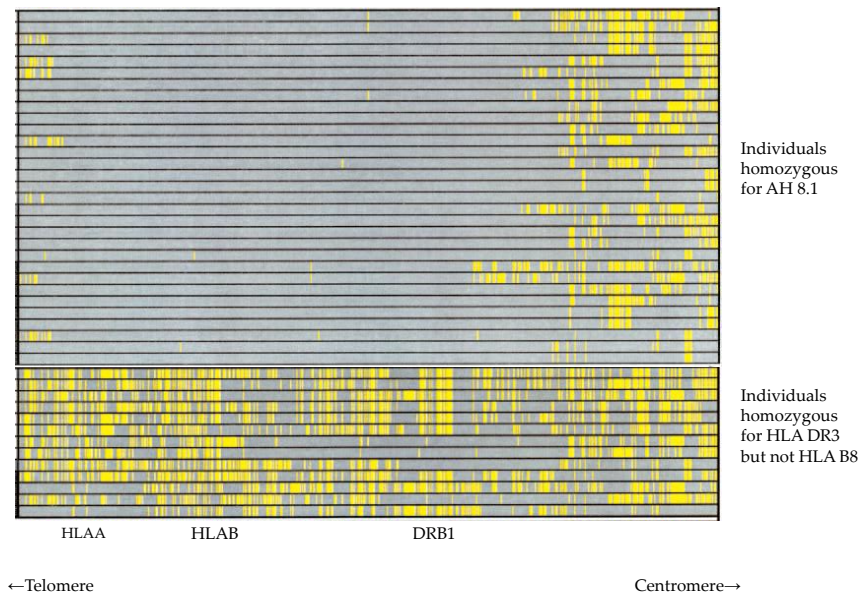


Figure 6. Alignment of 9 kb sequence at HLA-B. Sequences of 6 individuals with homozygous ancestral haplotypes were downloaded from UCSC browser [33] at HLA B and aligned using ClustalX2 [34]. For the purposes of illustration only, common sequences were removed and the interruption marked as //. The nucleotides of AH 44.2 are displayed in

the first row. Nucleotides of AH 62.1, 7.1, 44.1*, 8.1 and 18.2* are given only where they differ from AH44.2 and otherwise marked with a dot. Missing nucleotides are marked with a dash and shaded grey. The sequences are described by Horton et al. [24], whereas AH haplotypes have been assigned from the HLA allele types given by Horton, according to Cattley [35].

The degree of conservation of each ancestral haplotype is truly remarkable. For example, Smith et al. [32] found variation at only 11 of 3, 600, 000 positions between HLA-A and DR. Similar findings have been reported by others, including Aly et al. [31], see Figure 7. Mutation and recombination must be suppressed.

Figure 7 illustrates the importance of interpreting nucleotide diversity according to the block structure of the genome. Thus, conservation in the intervening, essentially monomorphic regions, is of minor interest, whereas differences within PFB allow the discovery of evolution, function and disease susceptibility.



Adapted from ref. [31].

Figure 7. Remarkable conservation within 8.1 haplotypes. A total of 656 SNPs spanning 4.8 Mb in the MHC region are depicted. The lower frequency allele (row) for each SNP along each haplotype column is highlighted in yellow. The top group depicts SNP results from 8.1 AH haplotypes ($n = 31$), the lower group are HLA-DR3, non-B8 haplotypes ($n = 13$). The 29.9 Mb range between HLA and DRB1 was >99.9% conserved, with only 9 variant alleles of the 10, 768 alleles identified for the 384 SNPs in the 31 8.1 AHs.

The inescapable conclusion is that some parts of the genome have *not* two or three but hundreds of alternative ancestral sequences.

6. Sequence analysis of ancestral haplotypes

The challenge in terms of sequence analysis is to compile a sufficient matrix to be able to recognize each haplotype and its extent. Assume access to multigenerational families with accurate, truly phased but unmolested raw sequences of at least 100, 000 bases:

1. Clustering of these by independent criteria relating to as many as hundreds of distinct ancestral haplotypes.
2. Alignments which take account of haplospecific duplicons, indels and retroviral-like elements (RLE).
3. Functional information to address biological and disease significance.

Given NGS, this approach is now feasible, even if daunting.

Importantly, those regions which are complex because of duplications and indels should be included rather than “corrected” based on the assumption that there is a single reference or “wild” sequence. Some examples are shown in Figure 6.

In designing better algorithms [36], the strategy for comparative analysis will be crucial. In many polymorphic regions, the density of differences can be as high as 1 per 10 bases when different haplotypes are compared but as low as 0 if the haplotypes are the same. It follows that analysis without haplotype assignment will be misleading.

7. Finding polymorphic frozen blocks and their ancestral haplotypes

The best clue to the location of these blocks is segmental duplication [17, 37].

To characterize the PFB, it is helpful to amplify haplospecific geometric elements [30], see also Table 3. Essentially, this approach reveals duplications as seen in Figure 8. McLure developed the approach to find PFB throughout the genome [36]. Paralogous regions are also helpful as shown in Figure 9.

Once identified, we recommend tracking the polymorphism through panels of multigenerational families as illustrated in Figure 10. Although the region is over 10 megabases, recombination was not found. The different haplotypes in the three breeds must have been conserved for at least hundreds of generations and mark differences in function such as the melting point of fat [37].

8. Applications to NGS and the 1000 genomes project

8.1. Mapping PFB from 1000 genomes data

Since it is known that PFB can be mapped by plotting diversity measurements (see Figure 3), we asked whether it would be possible to use data from the 1000 Genomes Project [39] in the same way.

Earlier work was based on haplotypes defined in multigenerational families. Initially, sequences of haplotypes were determined from Sanger sequencing of homozygous cell lines. In contrast, variations in 1000 genomes are determined from NGS for heterozygous and unrelated

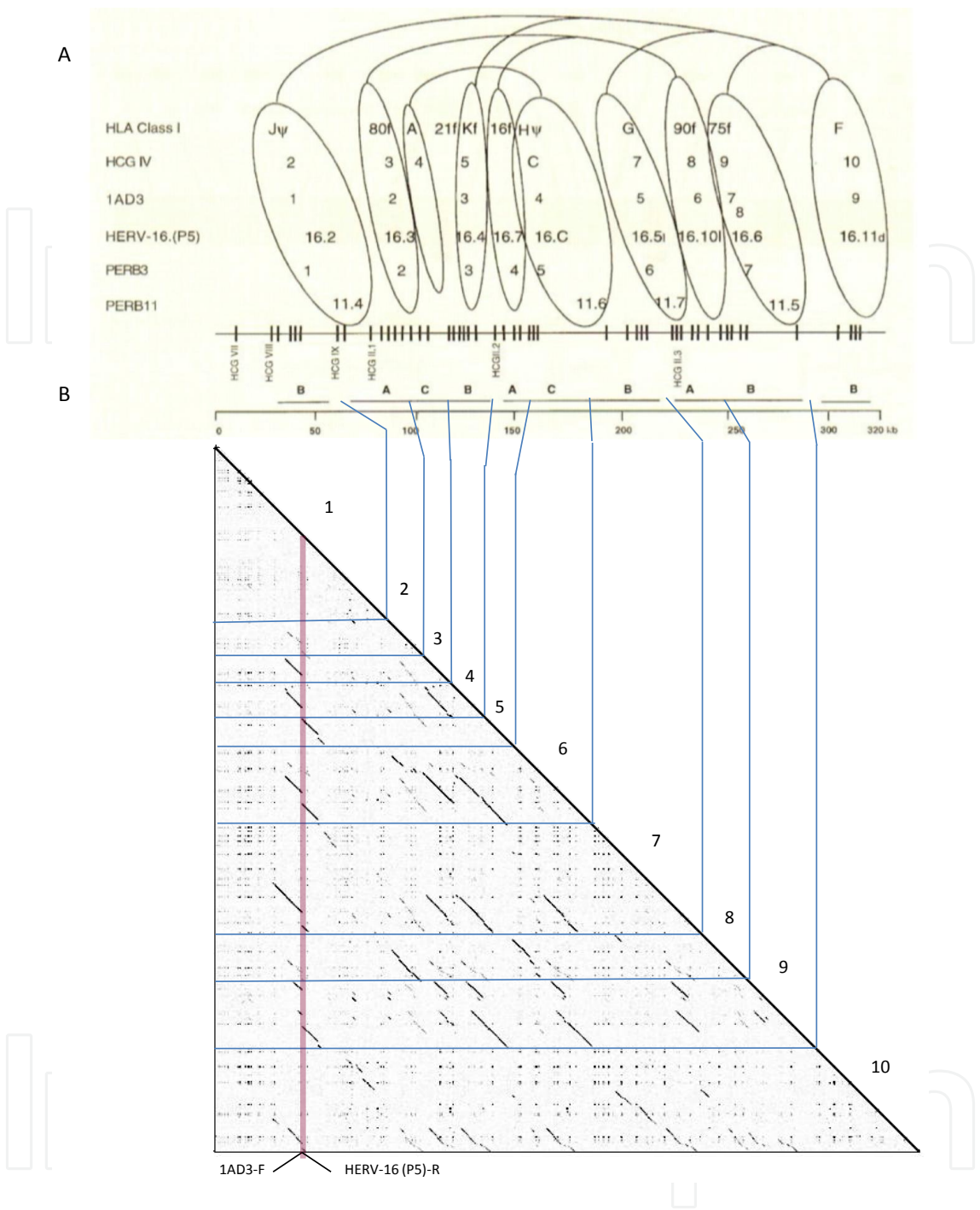


Figure 8. Segmental duplications in MHC alpha block. (a) Gene families and retroelements PERB 11, HLA, HCGIV, AD-3, HERV-16, PERB3 are duplicated to form an ordered pattern within the alpha block of the MHC, indicating that a segment containing multiple genes and retroelements has been duplicated to give 10 duplicons. Full-length duplicons consist of PERB11, HLA, HCGIV, 1AD3, HERV-16 (P5) and PERB3 genes. HLA-80, HLA-A, HLA-K, HLA-16, HLA-90 and HLA-F duplicons lack PERB11 gene. f = fragment, 1 = LTR only, d = discontinuous. ψ = pseudogene. A, B and C represent subgroups of duplicons with greater similarity. (b) A dot plot of the 319 kb genomic sequence encompassing the alpha block was compared against itself. The oblique lines in the plot represent duplications whereas the dots represent retroelements. Lines connect regions of the dotplot to the appropriate duplicons. The primers shown amplify products of different lengths in each duplication. Sequence from GenBank accession number AF055066. Adapted from ref. [17].

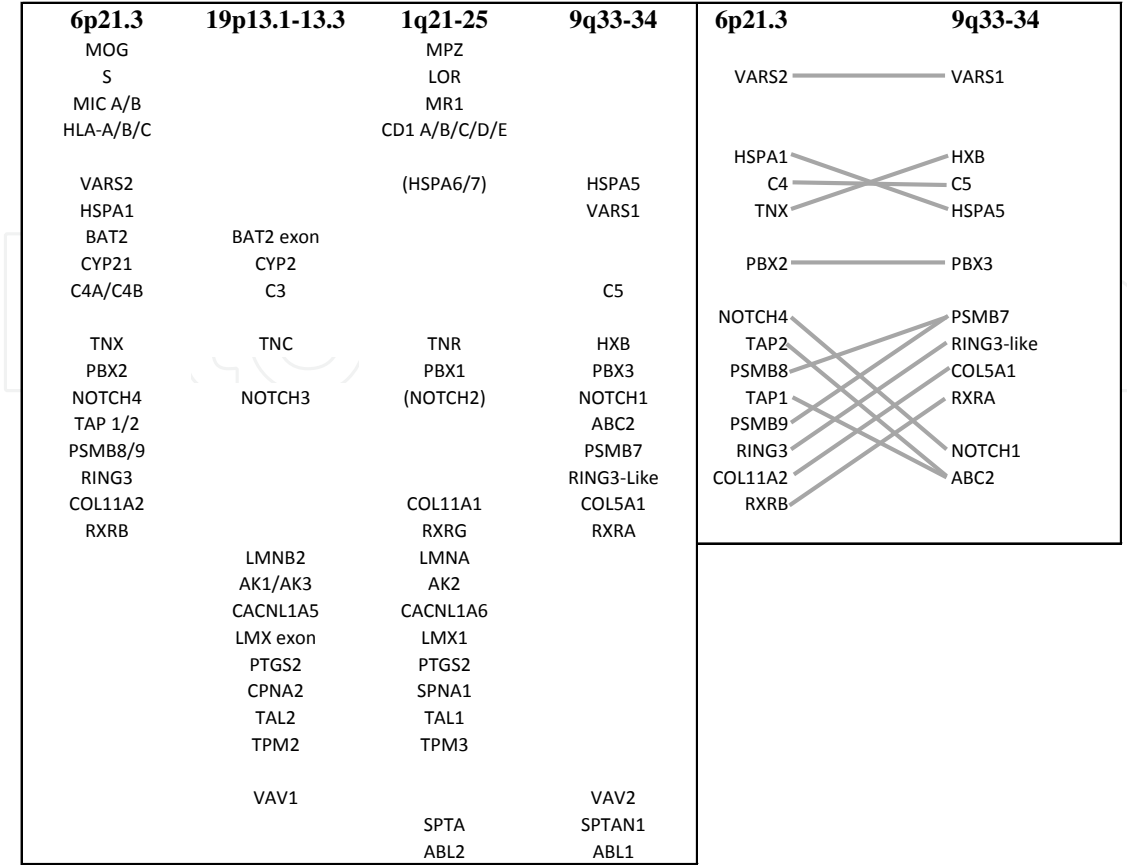
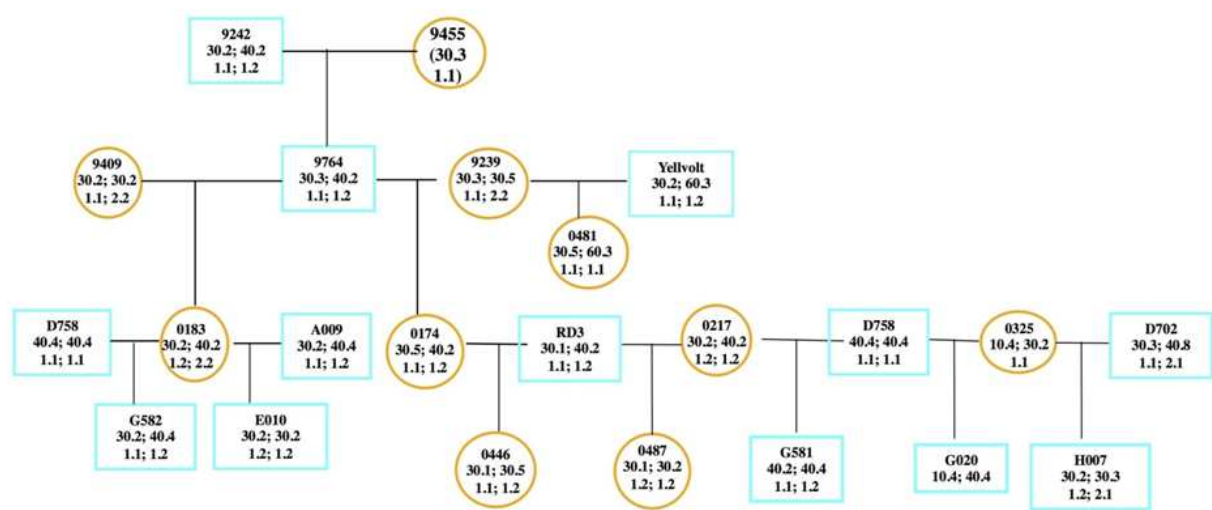


Figure 9. Paralogous locations of MHC genes. MHC genes are found on four chromosomes: 1, 9, 19 as well as chromosome 6. The arrangements of genes in each of the paralogous groups can be largely explained by duplication with and without inversion events. The genes common to chromosomes 6 and 9 are shown.

individuals. The phasing is an estimate based on ideas inherent in population genetics. It is known that the approach is a risky approximation. For example, artefactual “switch-overs” between haplotypes are misleading [40]. Since the reads tend to be short, such as just hundreds of bases, assembly can be fraught. There is a risk of missing complex polymorphisms and underestimating the number of ancestral haplotypes. Given these problems, we plotted several indices related to the 1000 genomes. The intention was to identify any similarities with the distribution as shown in Figure 3.

Unexpectedly, Figure 11 shows a remarkable correspondence between the classical measurements and our extraction from the 1000 Genomes database. The exception around 31.4 Mb was missed by the NGS reanalysis presumably because it is a region which is rich in complex iterative sequences, as shown in Figure 12.

These results are very encouraging in that the advantages of NGS can be coupled with identification of genomic architecture and therefore targeting of the most informative regions. The similarity, by simply counting the base differences per 10 kb, can be refined and applied to the whole genome. The plot of number of “haplotypes” is also promising, although clearly not indicative of the number of ancestral haplotypes.



Reproduced with permission from ref. [22].

Figure 10. Tracing segregation through three generation families. The alleles at MRIP, now known as myosin phosphatase Rho-interacting protein, are used to designate haplotypes within the 5.5 Mb region of bovine chromosome 19 from SREBF1 to TCAP. Within this region, there are many genes involved in muscle development, growth and fatty acid synthesis. For further details, see Williamson et al. [38].

8.2. Comparing polymorphic sequences of well-characterised PFB

Since there are numerous ancestral haplotypes within a PFB, it is essential to compare as many sequences as possible. An example is shown in Figure 6.

It can be seen that

- Only a minority of sites are informative and these must be selected from the remainder.
- Kilobases need to be examined and reduced 10- to 100-fold, retaining the informative sites.
- Different haplotypes are defined by specific combinations of bases at those informative sites.
- Very few single nucleotide polymorphisms are specific for a particular ancestral haplotype. On the contrary, specific combinations may be best defined by comparison with a library of reference sequences.
- Indels are important: alignments can be misleading.

Thus, although the identification of each of the many haplotype remains challenging, the overall patterns of informative sites are helpful in screening for PFB and for localising haplospecific sequences.

9. Conclusion

In analysing NGS databases, we recommend:

1. Screening for PFB.

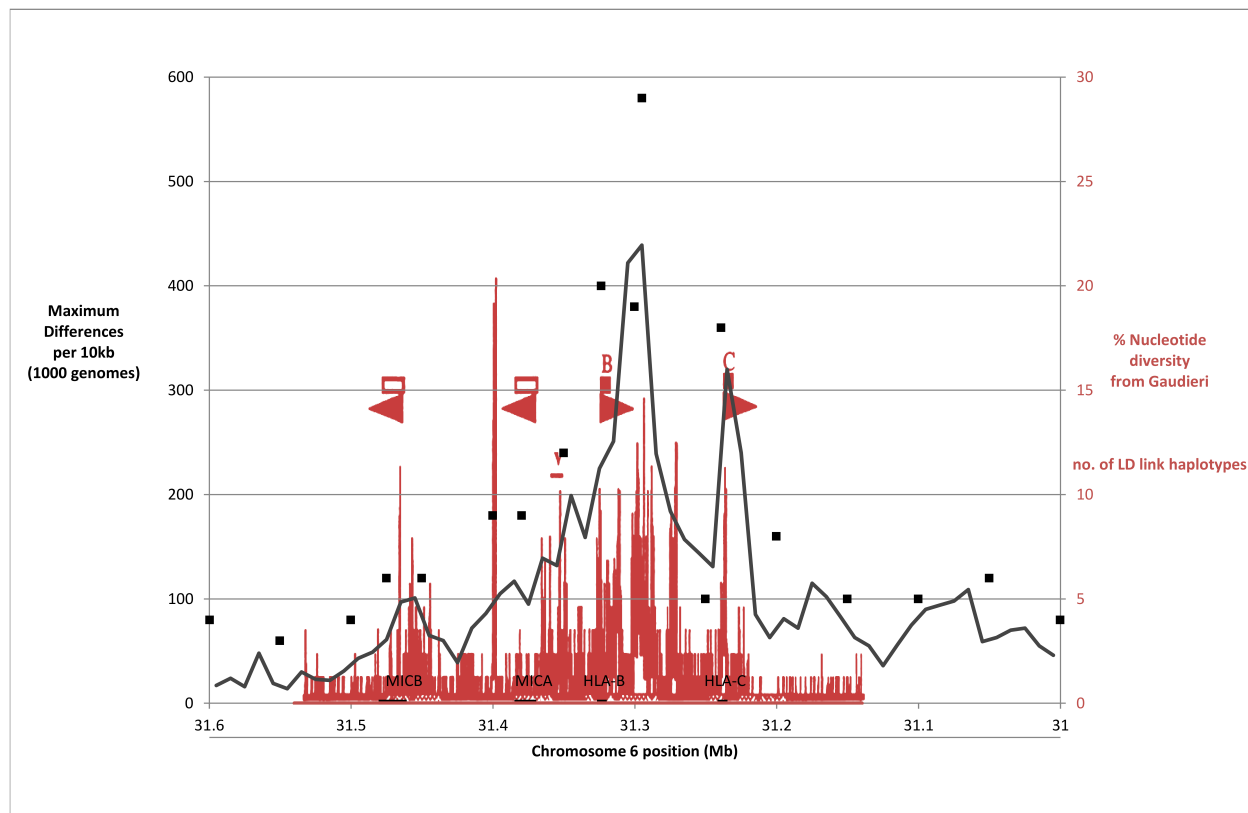


Figure 11. Regions of high sequence diversity within 1000 genomes are similar to previously identified PFB. Imputed haplotypes in the 600 kb region surrounding HLA-B from 553 individuals were downloaded from the 1000 Genomes browser [41]. The population groups chosen were of African, European and Asian origin (ACB, ASW, BEB, CEU, CHB and YRI). The majority of variations recorded in the 1000 Genomes vcf files are SNPs, but some indels up to 174 bp are recorded. For each imputed haplotype, we counted the number of differences from the reference sequence in 10 kb sections. Indels were counted as one difference, irrespective of length. The black curve represents the maximum difference at each 10 kb. The red lines, taken from ref. [42], show the amount of nucleotide diversity between two individual haplotypes, counted in 100 bp sections. Haplotypes compared for this section were 44.1 to 62.1, 44.1 to 8.1 and 8.1 to 14.1. Squares show the number of LD_link [41] “haplotypes”, calculated from sets of adjacent variants in 500 bp intervals. LD link requires that variants be biallelic and only takes single nucleotide changes, not indels. Only variants with at least two examples in the CEU and YRI populations were included.

2. Alignment based on the ability to detect multiple, and even hundreds of ancestral haplotypes.
3. Analysis must recognise that haplospecificity is confirmed by many characteristics including RLE, indels, copy number and complex iterative sequences.
4. Analysis may be facilitated by examining paralogous regions which help to define interactions, including epistasis.
5. Validation of results by showing segregation in multigenerational family studies.
6. Confirming biological significance by demonstrating permissive or *sine qua non* associations.

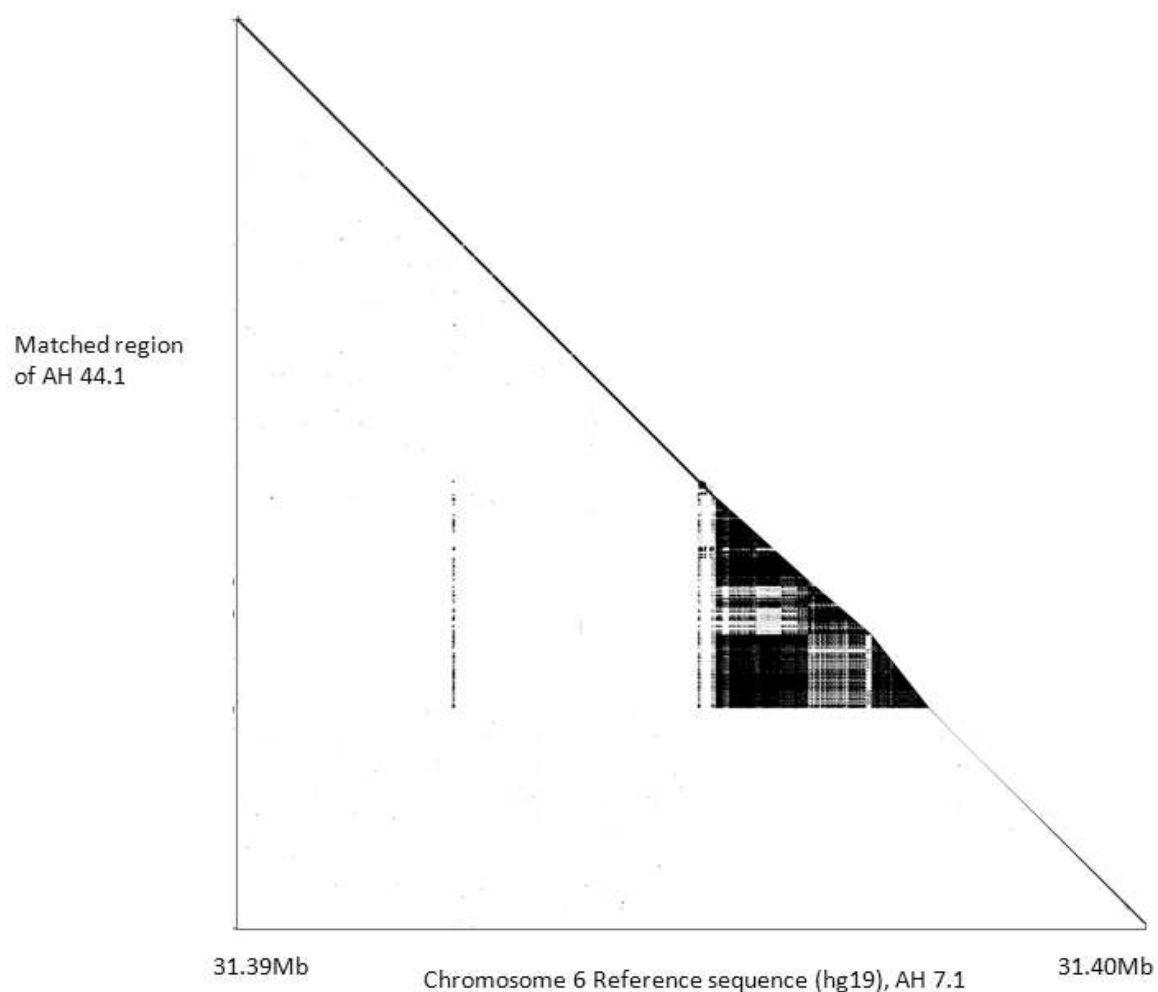


Figure 12. Complex iterative element. Dotplot of a 10 kb region in the MHC between MICA and MICB showing a complex iterative element. Gaudieri [42] shows high nucleotide diversity for this region which was not recorded within 1000 Genomes data. Example sequences for AH 7.1 and AH 44.1 downloaded from UCSC genome browser. Dotplot generated with Gepard [43] using word length 10.

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